
Validation of Integrative Models

Advanced Analysis Methods

IMP workshop

Dec. 16, 2016

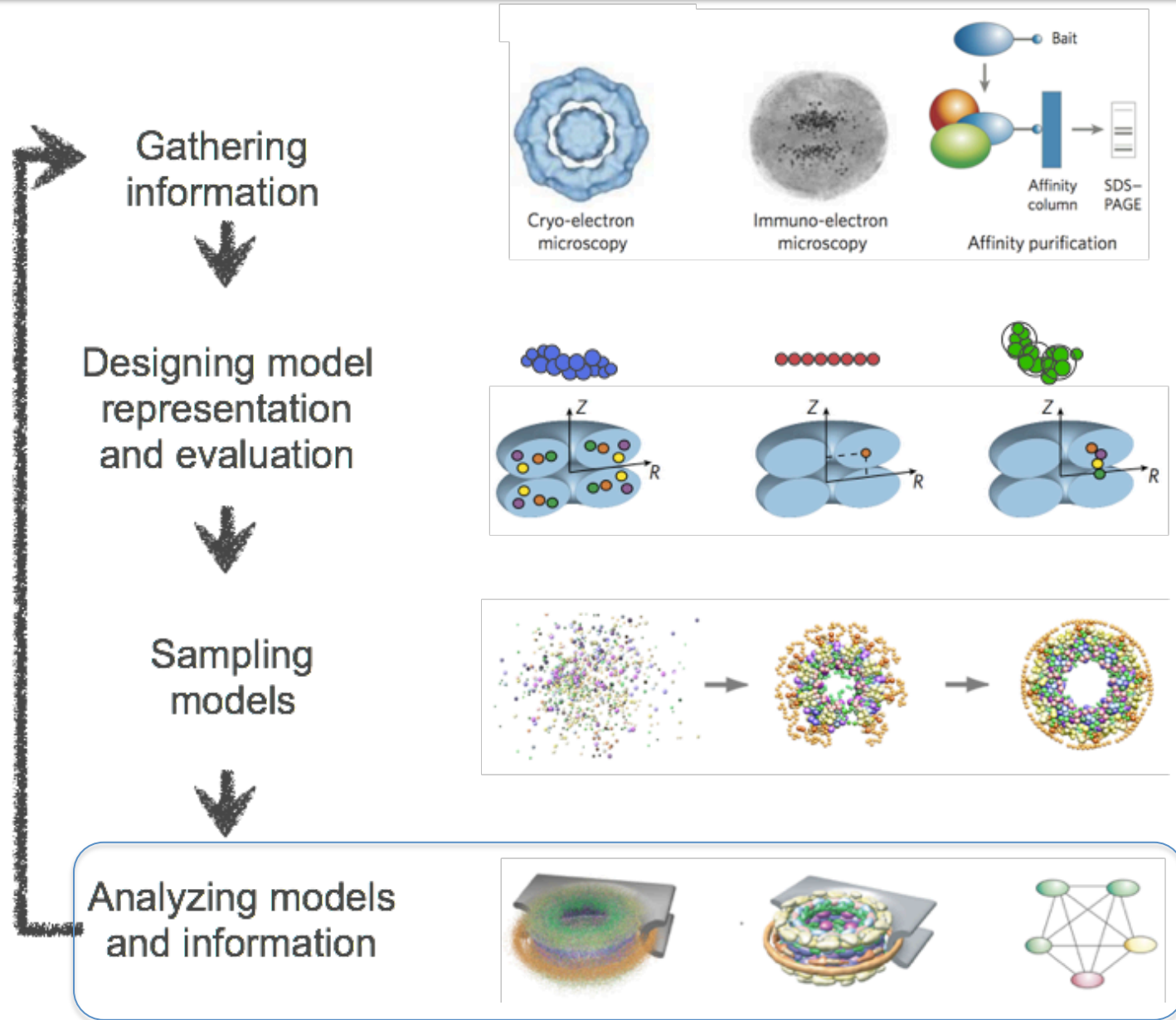
Daniel Saltzberg

saltzberg@salilab.org

Shruthi Viswanath

shruthi@salilab.org

Four Stages of Modeling



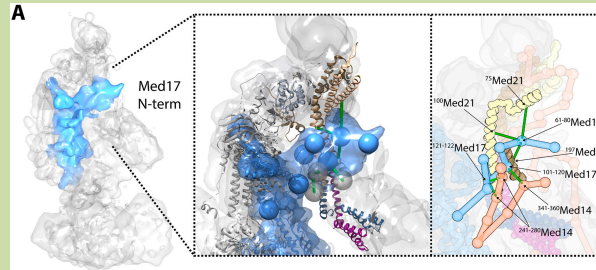
Outcomes of structural modeling

Many models are wrong.
Some models are useful. -Andrej Sali

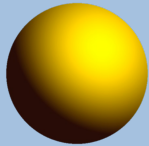


Useful Model!

Robinson, Trnka et. al. 2015. eLife

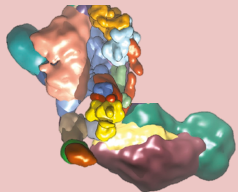
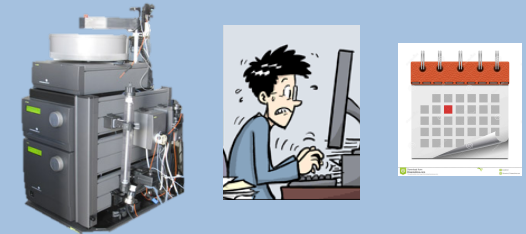


Biological insight!



Unuseful Model

~\ (ツ) ~\



Wrong Model

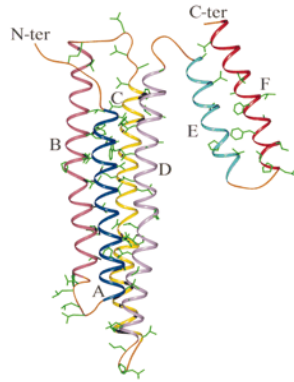
Incorrect claims



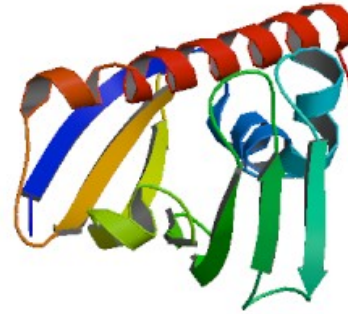
* The IMP developers make no guarantees of Nobel prizes based on use of the software

Yes, there are bad models...

Fraud



Apolipoprotein A1
(2005)



Birch Pollen Allergen
(2010)

Mistakes

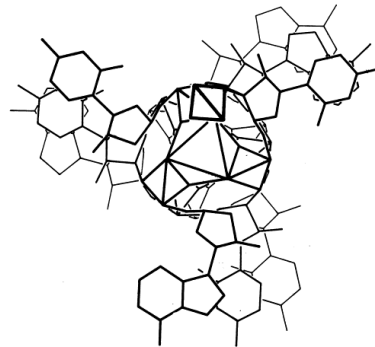
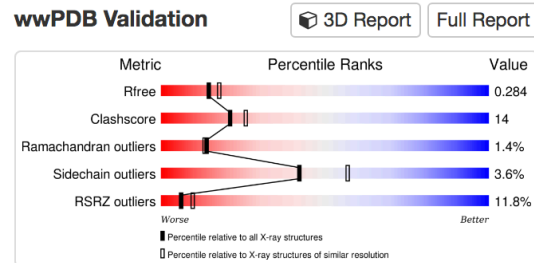


FIGURE 6
Plan of the nucleic acid structure, showing several nucleotide residues.

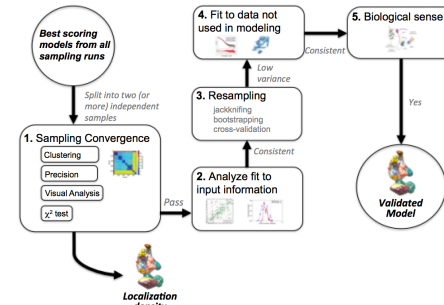
Nucleic Acid
(1953)

Validating / interpreting integrative models

■ Methodology under development



Crystallography validation protocols are fairly well established



This workflow is current as of last week

- Complex analysis in IMP requires customized scripts
 - We're developing pipelines to perform these methods

A subset of where can modeling go wrong

Gathering
information

Incorrect
info

Bad data

Experimental
inconsistencies

Bad
homology
models

Incorrect
assumptions

Designing model
representation
and evaluation

Poorly
defined
restraints

Representation not
commensurate
with data

Overfitting

Sampling
models

Insufficient
sampling

Miss global minimum

Miss important state

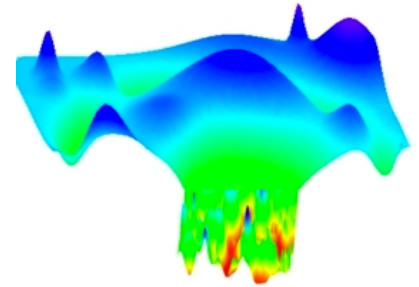
Analyzing models
and information

Model does
not satisfy
information

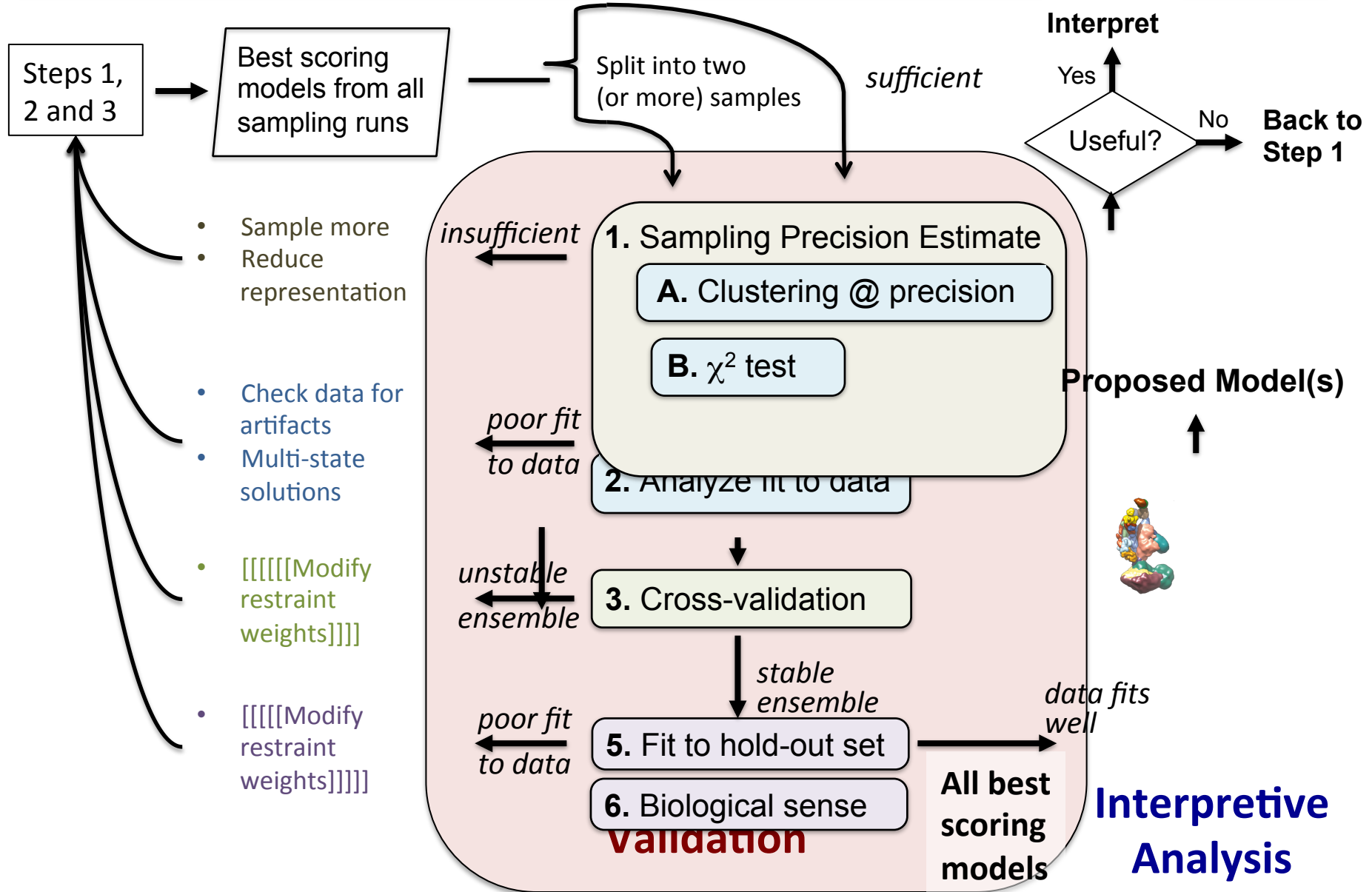
Reporting too
high of a
precision

What to validate?

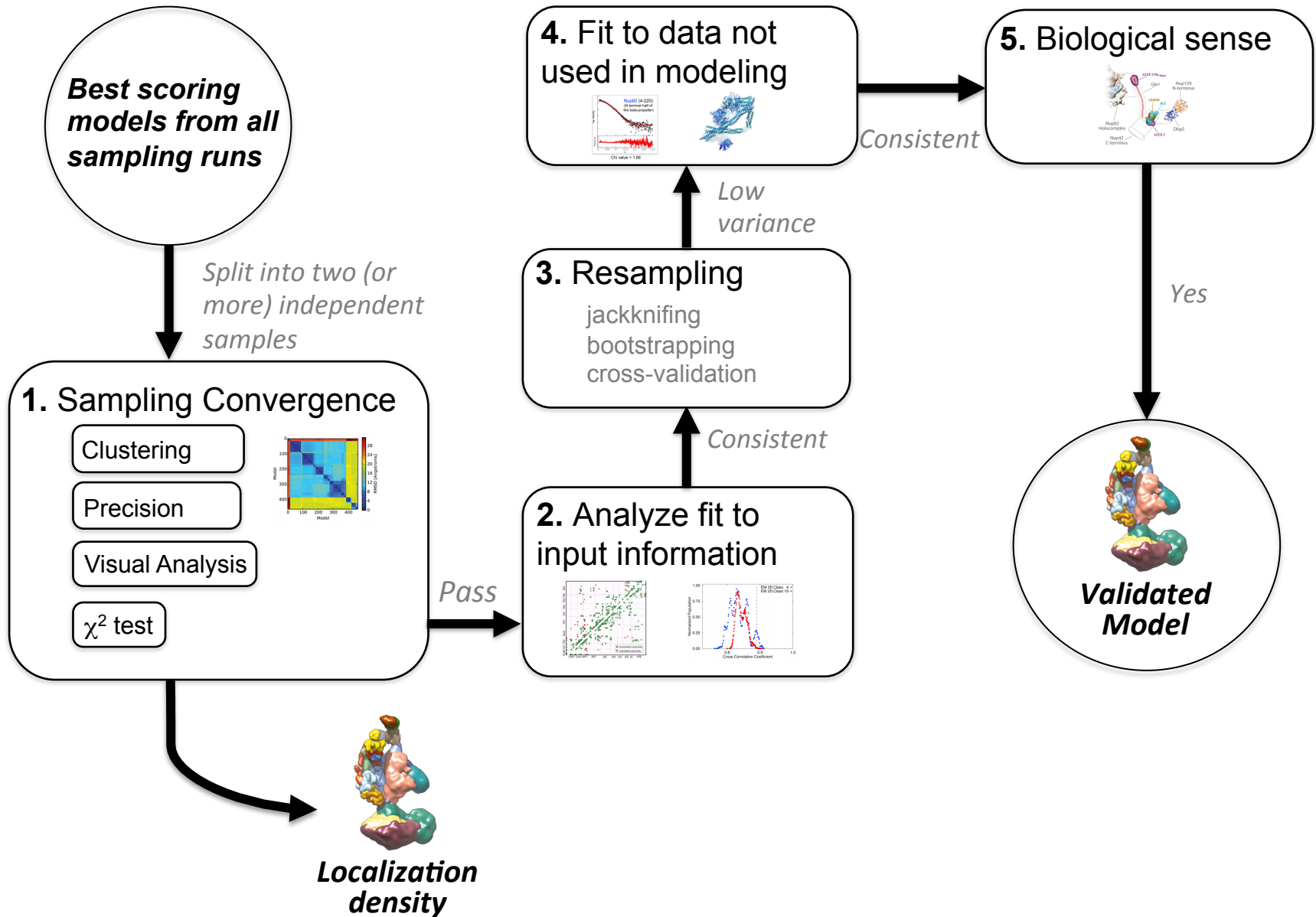
- **Sampling Exhaustiveness**
 - Possible sampling missed a subset of good scoring models
- **Fit to Data/Restrains**
 - Poorly fit data may indicate problem with data/modeling
- **Jackknifing**
 - Guard against overfitting
 - Complete cross-validation
 - Like a composite omit map
- **Validation against other data**
- **How to proceed:**
 - All models



Step 4: Practical Analysis Flowchart

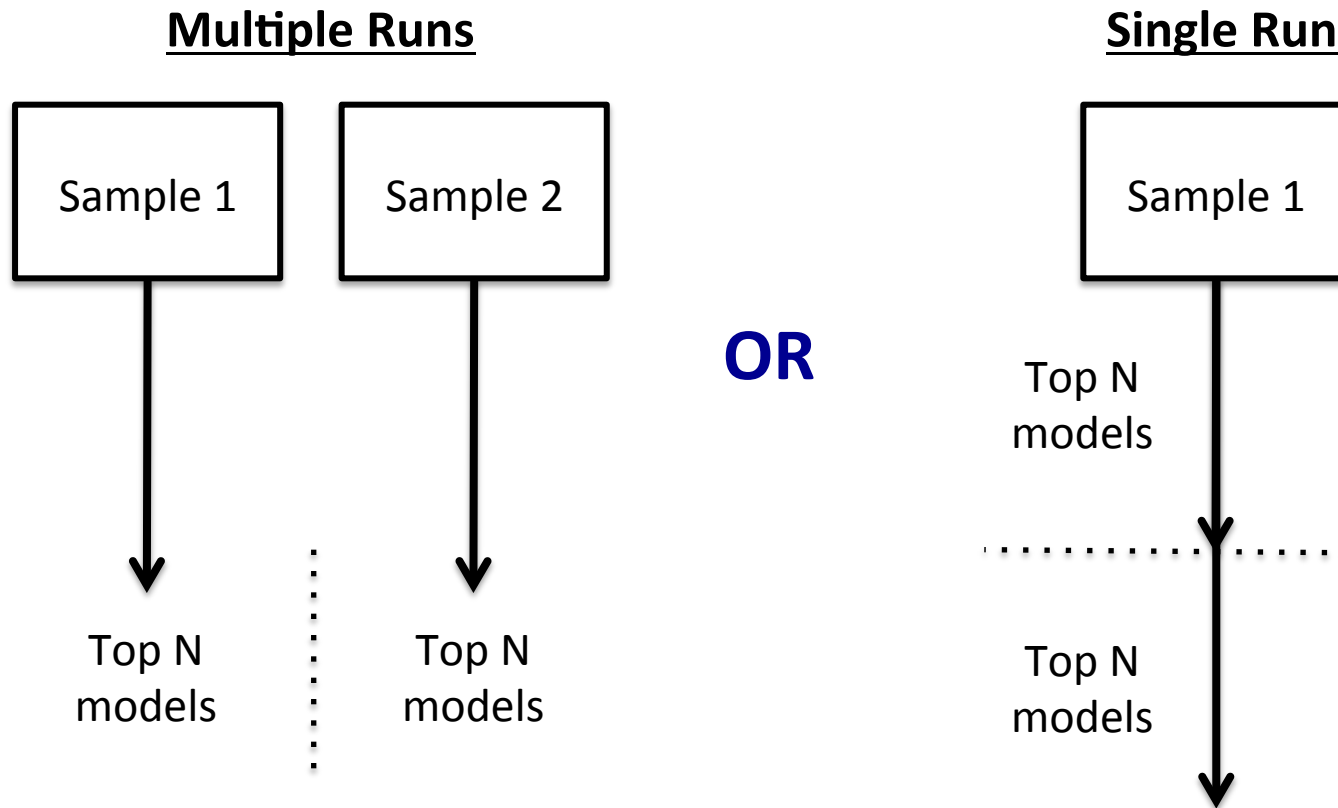


Step 4: Analysis



0. Pre-processing

- Split sampling into multiple independent sets

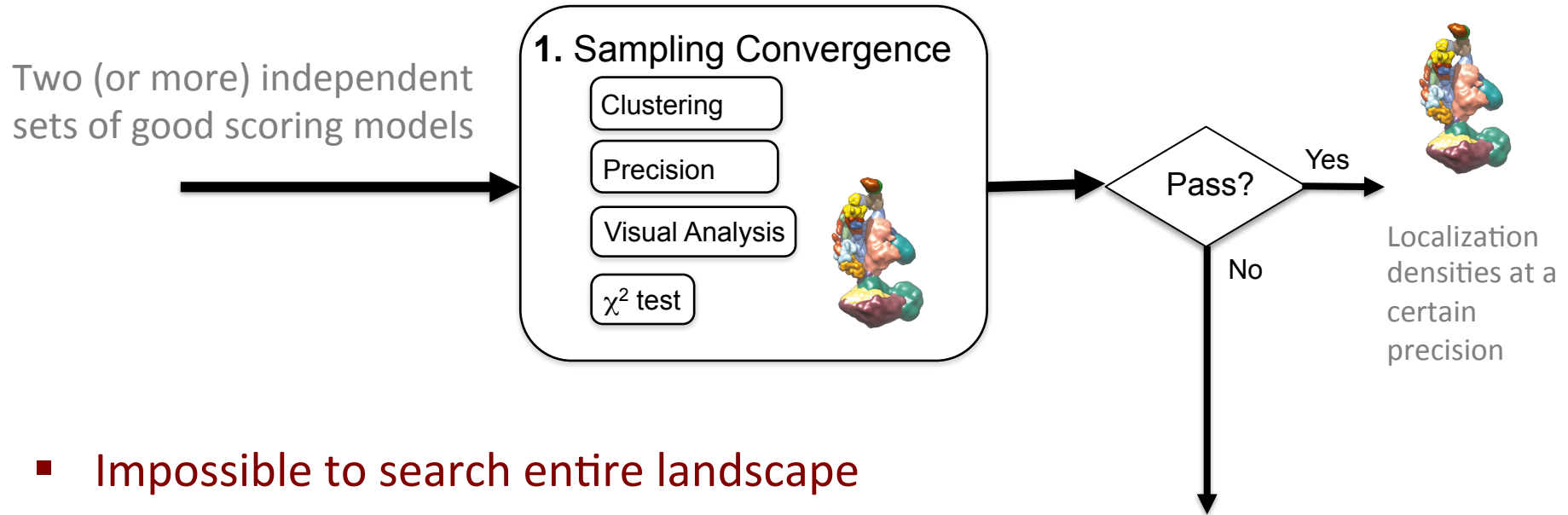


0. Pre-processing

- **Split sampling into multiple independent sets**
- **Gather best scoring models**

[illegible]

1. Assessing Sampling Exhaustiveness



- Impossible to search entire landscape
- **Method:** Compare independent samples of models
 - **Visual analysis:**
Compare localization densities.
 - **Statistical (in)significance:**
Show no statistically significant differences between clustering results

- **Sample more**
- **Reduce sampling space**
 - add more information
- **Reduce DOF**
 - reduce representation
 - impose symmetry

** No method gives proof of convergence*

1. Assessing Sampling Exhaustiveness

■ Visual Analysis

- Get clusters and localization densities for each independent cluster

```
import IMP
import IMP.pmi
import IMP.pmi.macros

rmf_dir = ./rmfs/      # path to the rmf directory
num_rmfs = 4           # number of rmfs in the directory
num_clusters = 1

# Setup macro
model = IMP.Model()
mc = IMP.pmi.macros.AnalysisReplicaExchange0(model)

rmsdc = {"B":"B"} # compo
alignc = None

densityc = {"Spc97":["Spc97"],"Spc98":["Spc98"],"Tub4":
["Tub4"],"Spc110":["Spc110"]}
#densityc = None

mc.clustering (rmsd_calculation_components=rmsdc,
               number_of_clusters=num_clusters,
               display_plot=True,
               number_of_best_scoring_models=num_rmfs,
               exit_after_display = False,
               rmfsdir=rmf_dir,
               density_custom_ranges = densityc)
```

Yeast Mediator Complex

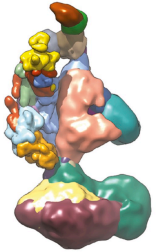
Total ensemble
of solutions



First half
ensemble



Second half
ensemble

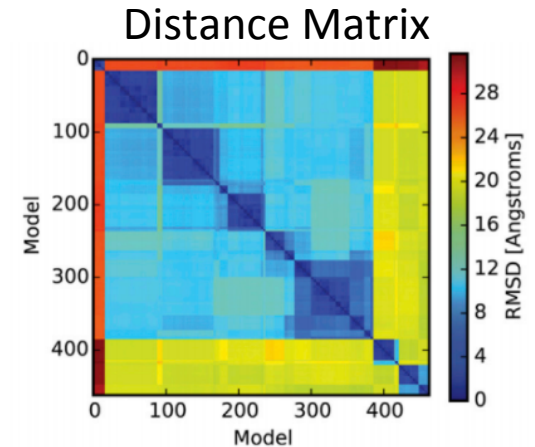


Robinson, Trnka et. al. 2015. eLife

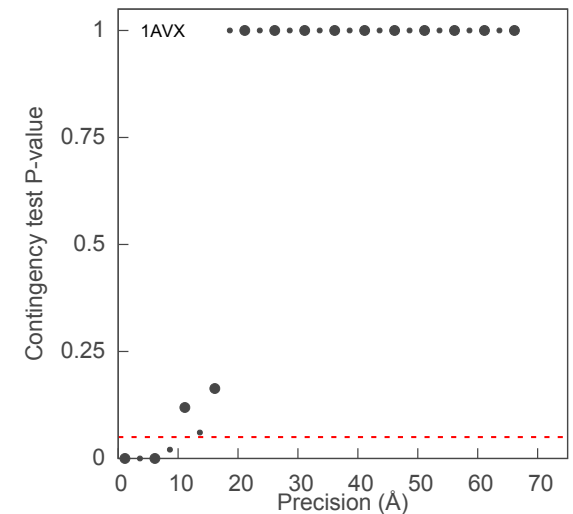
1. Assessing Sampling Exhaustiveness

■ Clustering and Precision

- Distance matrix is determined by pairwise C_{α} RMSD calculation
- **k-means** is used to separate into clusters based on RMSD
 - Must specify the number of clusters
- How many clusters to choose?
 - Visual analysis
 - Clustering metrics
- Clustering choices determine precision of your models
 - Many clusters – high precision
 - Fewer clusters – low precision

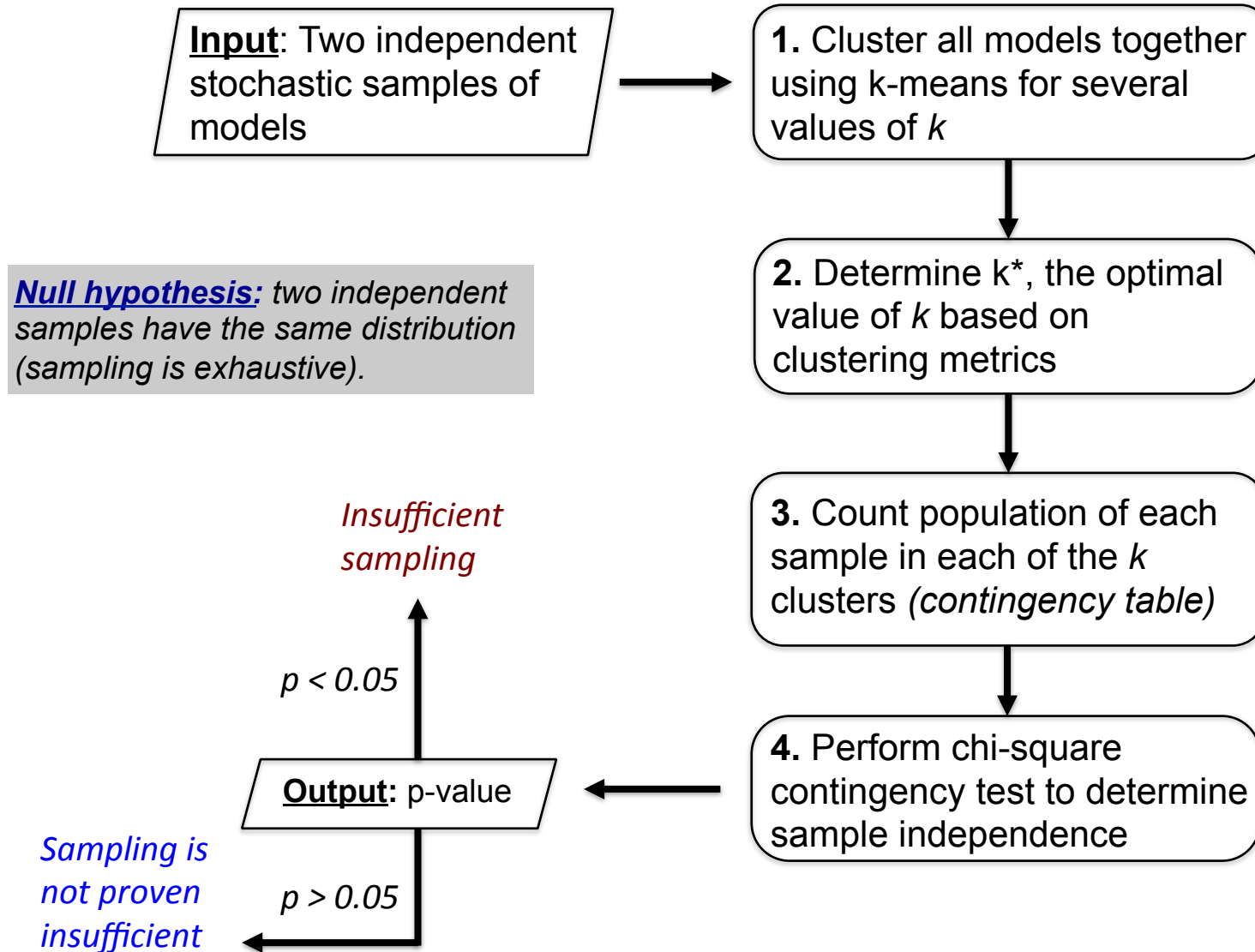


Nup82 – Top 463 models



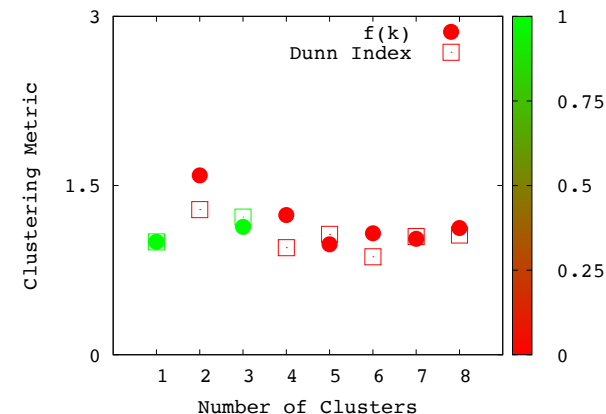
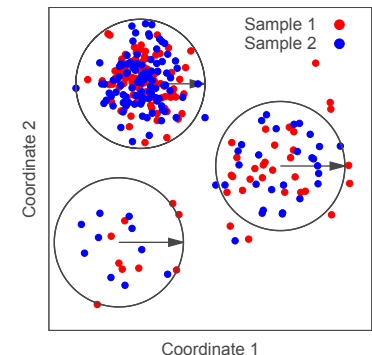
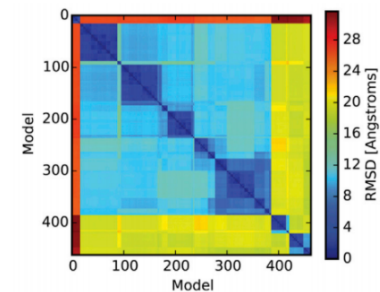
1. Assessing Sampling Exhaustiveness

Chi² Sampling Test Flowchart



Chi-squared convergence test

- **INPUT:** Get N top scoring models for each run from the output of sampling
`get_top_models_each_run.py <N>`
- **1. Clustering:** Perform k-means clustering on the combined set of models
`cluster_kn.py`
`precision_rmsf.py`
- **2. Determine k^* :** Determine the optimal value of k using clustering metrics
`metric_wrapper.sh`
 - **Dunn Index:** ratio of minimum inter cluster precision to maximum intra cluster precision.
`metric_dunn.py`
 - **Distortion Index:** $f(k)$: does having k clusters produce a smaller distortion than having $k-1$ clusters?
`metric_fk.py`



Contingency table and p-value calculation

- **3. Population Count:** Calculate number of models from each run in all clusters to form *contingency table*

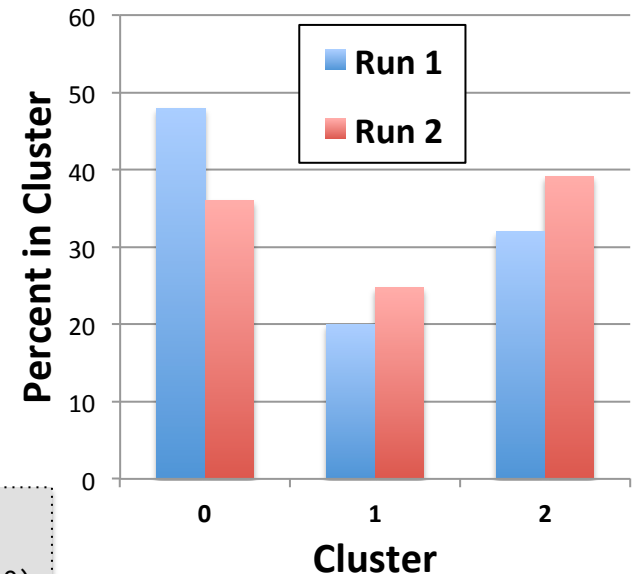
`get_models_per_cluster_kmeans.py`

- **4. Calculate *p*-value:** A $p\text{-value} < 0.05$ indicates a statistically significant difference between populations and incomplete sampling

`test_sampling_convergence.py`

```
numModelsFile = sys.argv[1] # file with number of models per cluster
modelsArray = numpy.loadtxt(numModelsFile)
percentArray = numpy.transpose((modelsArray/modelsArray.sum(axis=0)) * 100.0)
[chisquare,pvalue,dof,expected]=scipy.stats.chi2_contingency(percentArray)
print "P-value",pvalue
```

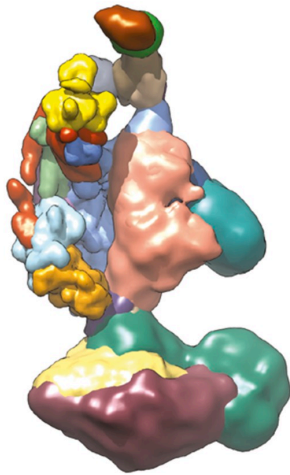
	Pct. of Run in Cluster	
Cluster	Run 1	Run 2
0	48.0	36.0
1	20.0	24.8
2	32.0	39.2



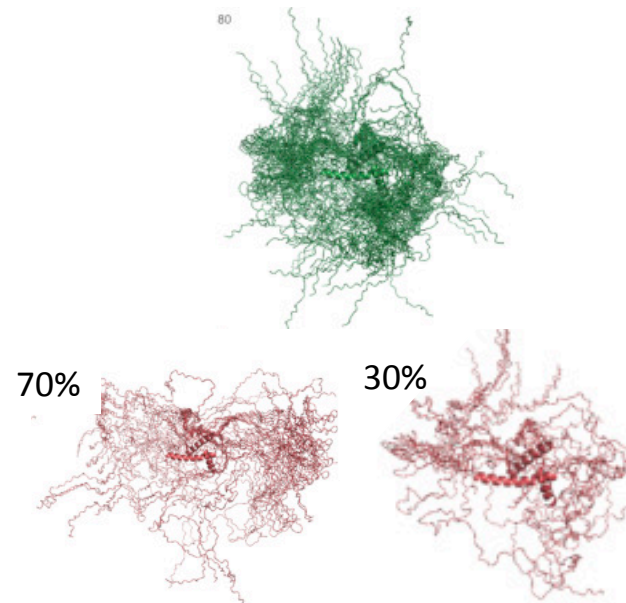
p-value = 0.228

1. Assessing Sampling Exhaustiveness

- **Output:**
 - Clusters
 - Localization Density (or Ensemble)
 - Precision

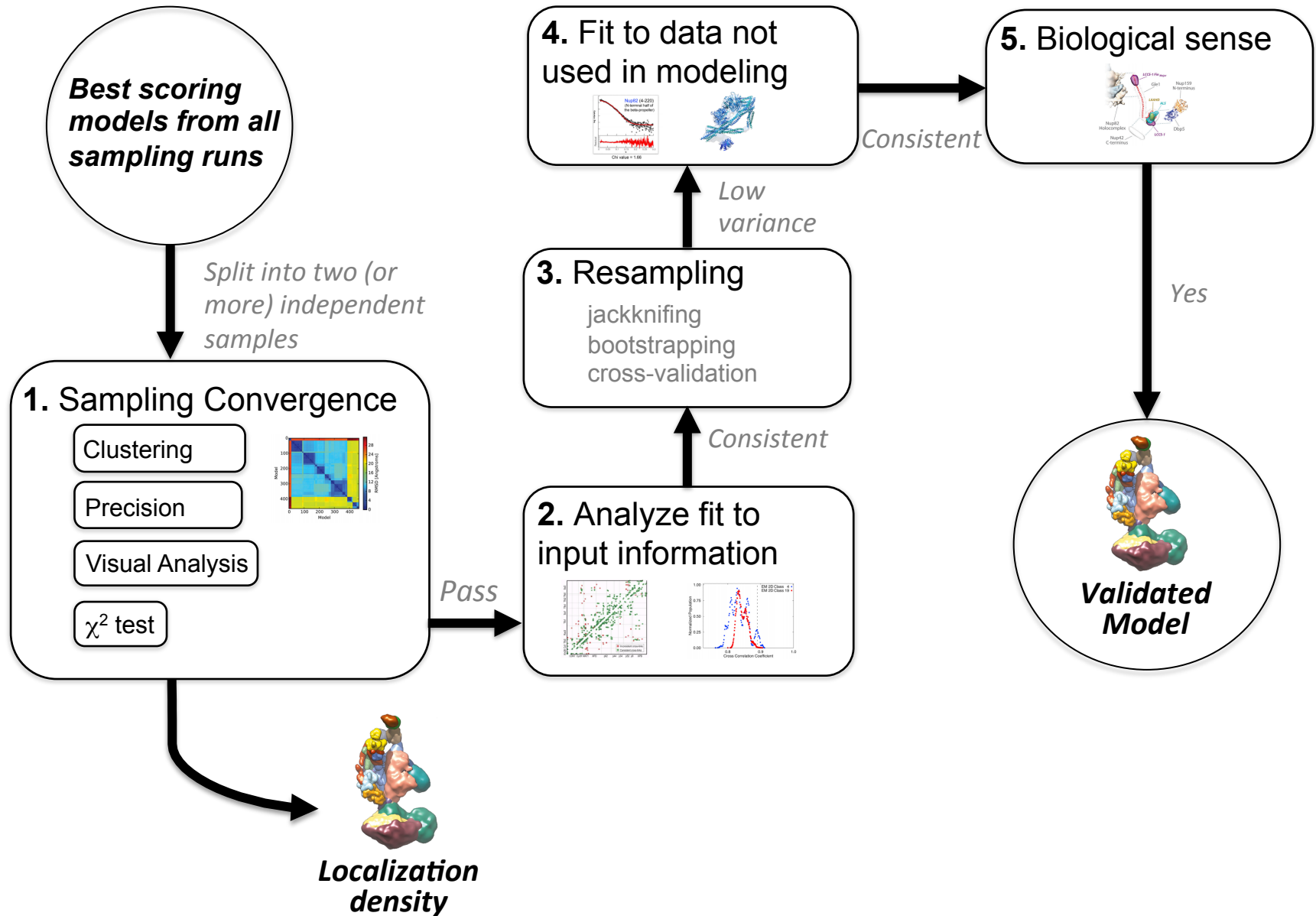


Single cluster ensemble

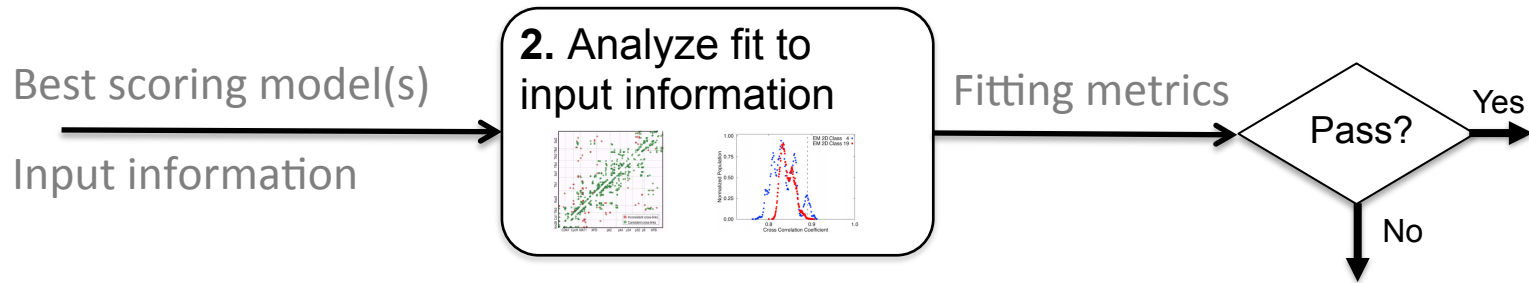


*Comparison of **single** and **multi-state** ensembles*

Step 4: Analysis



2. Assessing Fit to Data



- **Method:** Does the resulting ensemble of best scoring models actually represent the input data?
 - Passing criteria are subjective
- **Examine restraints that are not satisfied by any model**
 - Artifacts
 - Different experimental conditions
- **Evaluate a multi-state model**
 - Can you satisfy the model with two states simultaneously

2. Assessing Fit to Data

▪ Assessing Violations by Data Type

▪ Crosslinks

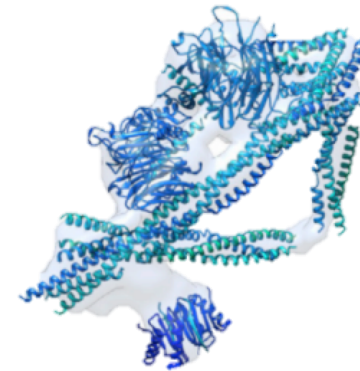
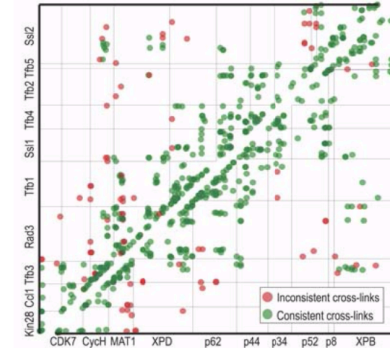
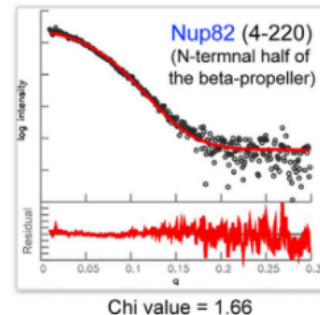
- Distance violations
- Score violations

▪ SAXS

- χ^2 value
- Radius of Gyration

▪ EM

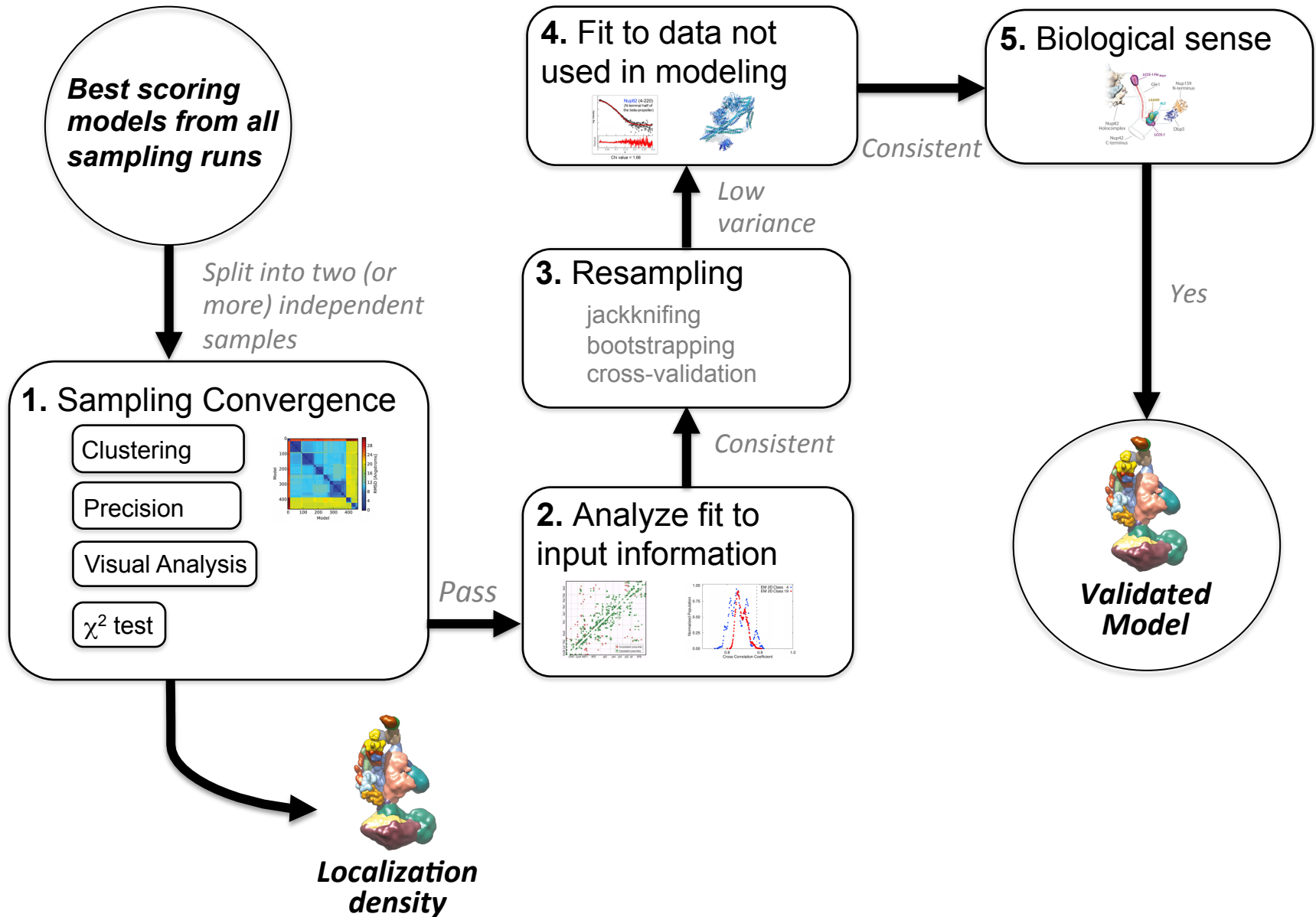
- Cross Correlation
- Visual inspection



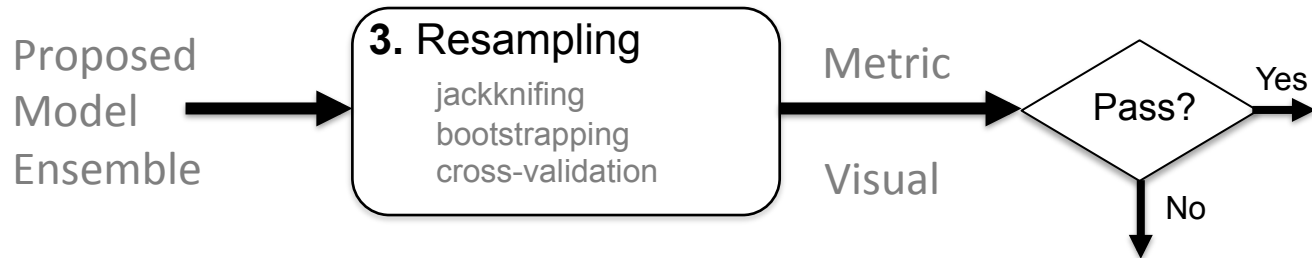
Subjective Questions:

- How do we define a violation?
- How many violations define a failing model?

Step 4: Analysis



3. Resampling Methods



- **Recalculate models using subsets of the data**

- **Bootstrapping**

- Remove random subsets of data

- **Jackknifing**

- Remove systematic subsets of data

- **Cross-validation**

- Predict values of held-out data
 - Score to original data

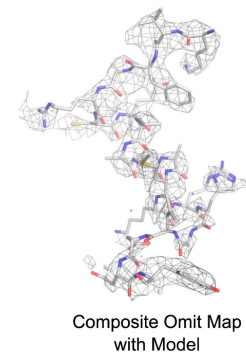
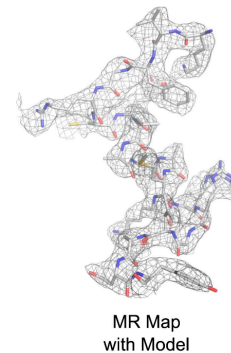
- **Prevent overfitting to certain data**

- **Assess the stability of the model ensemble with respect to target data.**

- **Model is too dependent on certain data**

- Reduce weight of the offending data

- **Data is not self-consistent**



Similar to calculating the composite omit map

3. Resampling Methods

■ Jackknifing

■ Omit pieces of data

■ Whole sets

- EM
- SAXS

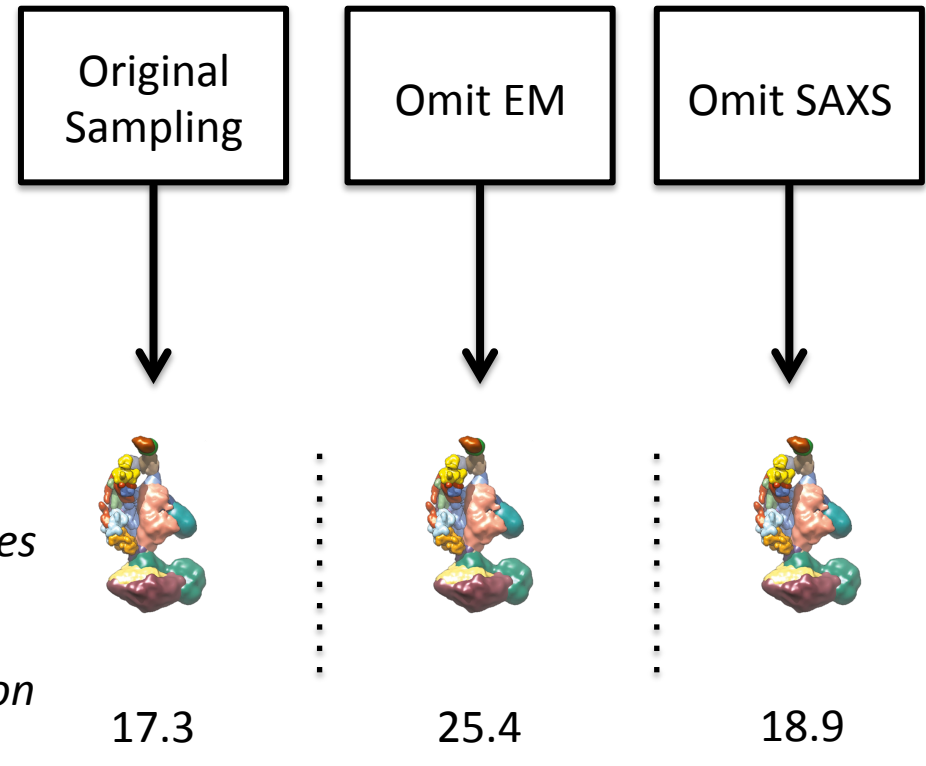
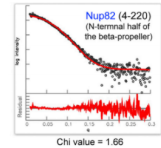
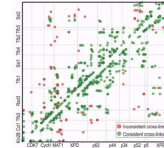
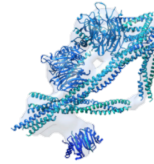
■ Subsets

- XL

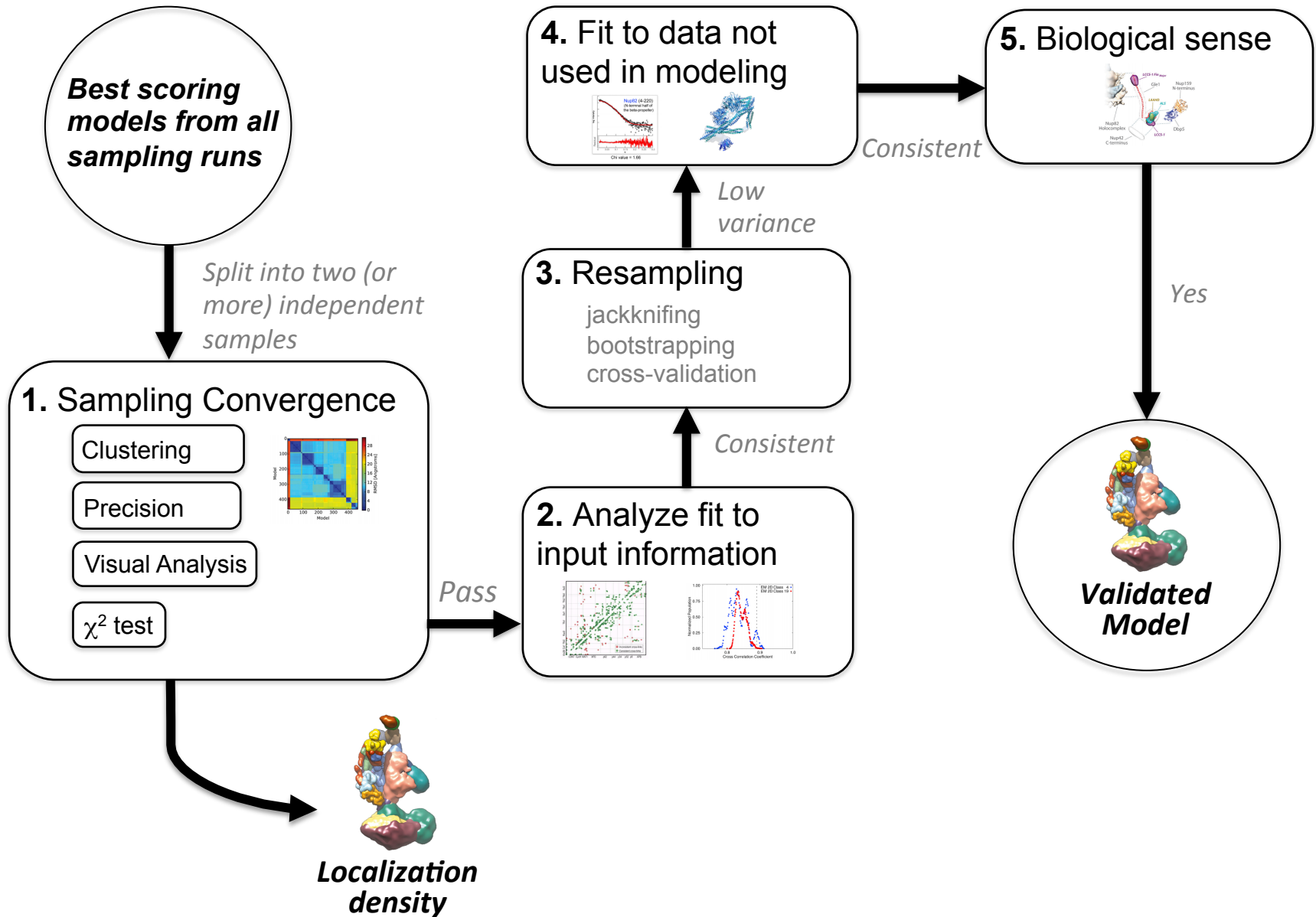
■ Densities similar?

■ Precision similar?

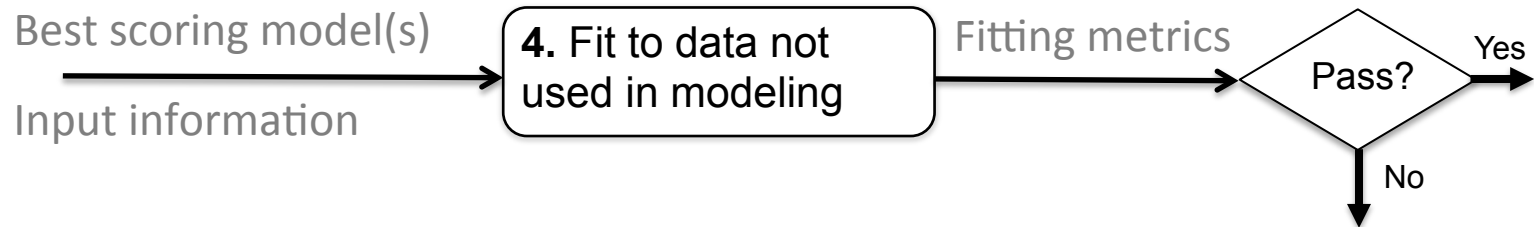
Practical Considerations:
Recalculating the entire ensemble is expensive.



Step 4: Analysis



4. Fit to Information Not Used in Modeling

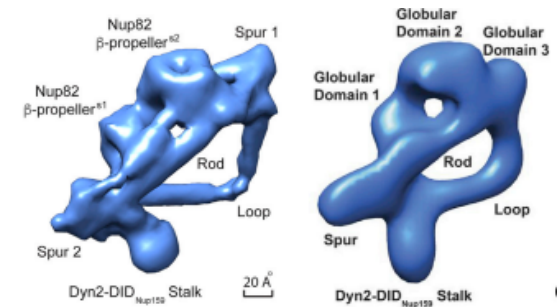


- **Same methodology as Step 2**

- Pre-defined hold-out set
- Information that is difficult to embed in a restraint
- Information from a slightly different construct
- New information collected after modeling

- **Examine restraints that are not satisfied by any model**

- Artifacts
- Different experimental conditions



Fernandez-Martinez et al. 2016

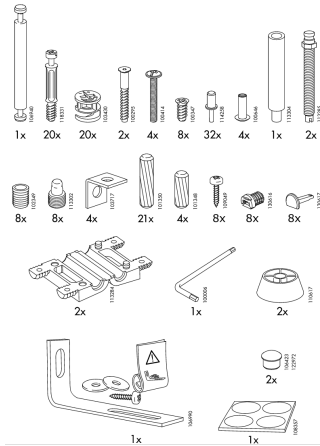
Gaik et al. 2015

Comparison of Nup82 models to negative stain EM of truncated model

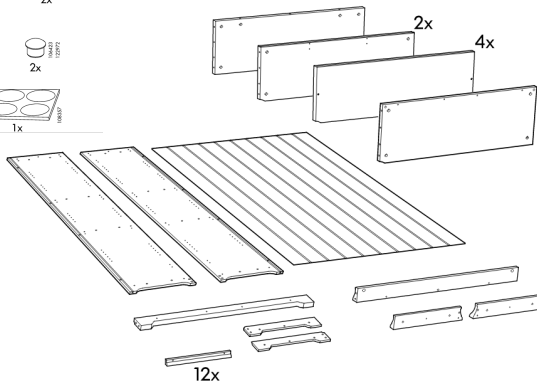
5. Biological Significance

- **The utility of the model is, in itself, a validation.**

- Satisfaction of patterns unlikely to occur by chance
- A wrong model is not likely to make sense



Supposed to be a bookshelf



- Poor book holder
- Pretty unstable

Probably incorrect



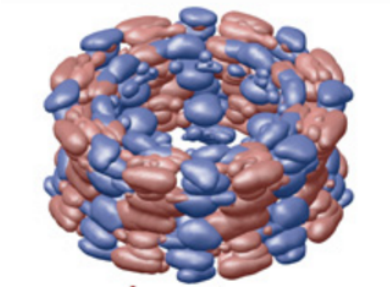
- Can hold books
- Looks like IKEA
- Doesn't fall apart

Probably correct

5. Biological Significance

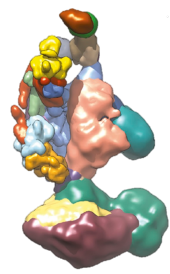
- **The utility of the model is, in itself, a validation.**
 - Satisfaction of patterns unlikely to occur by chance

Observation of suspected 16-fold
symmetry in the NPC

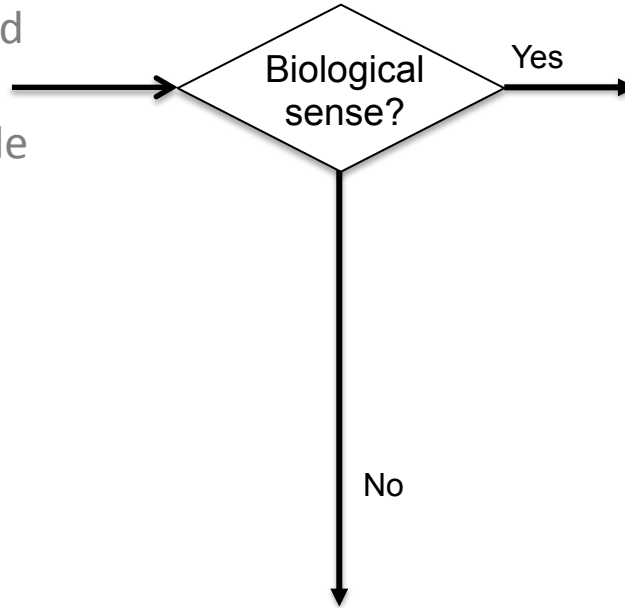


Alber, Frank, et al. "The molecular architecture of the nuclear pore complex." *Nature* 450.7170 (2007): 695-701.

5. Biological Significance



Proposed
Model
Ensemble



Reasonable confidence that model
is correct



Model is not necessarily wrong,
but care must be taken in any
new claims

What if I need more information?

- **Look outside of traditional structural biophysical experiments**
 - CoIP
 - Hydrogen/Deuterium Exchange
- **Make simple assumptions**
 - Symmetry
 - Interface
 - Oligomerization states
 - Stoichiometry

Communicating model validation

▪ Recent examples

Fernandez-Martinez et al., Structure and Function of the Nuclear Pore Complex Cytoplasmic mRNA Export Platform, Cell (2016), <http://dx.doi.org/10.1016/j.cell.2016.10.028>

Robinson, Philip J., et al. Molecular architecture of the yeast Mediator complex, Elife 4 (2015), <http://dx.doi.org/10.7554/elife.08719>

Integration into the WWPDB

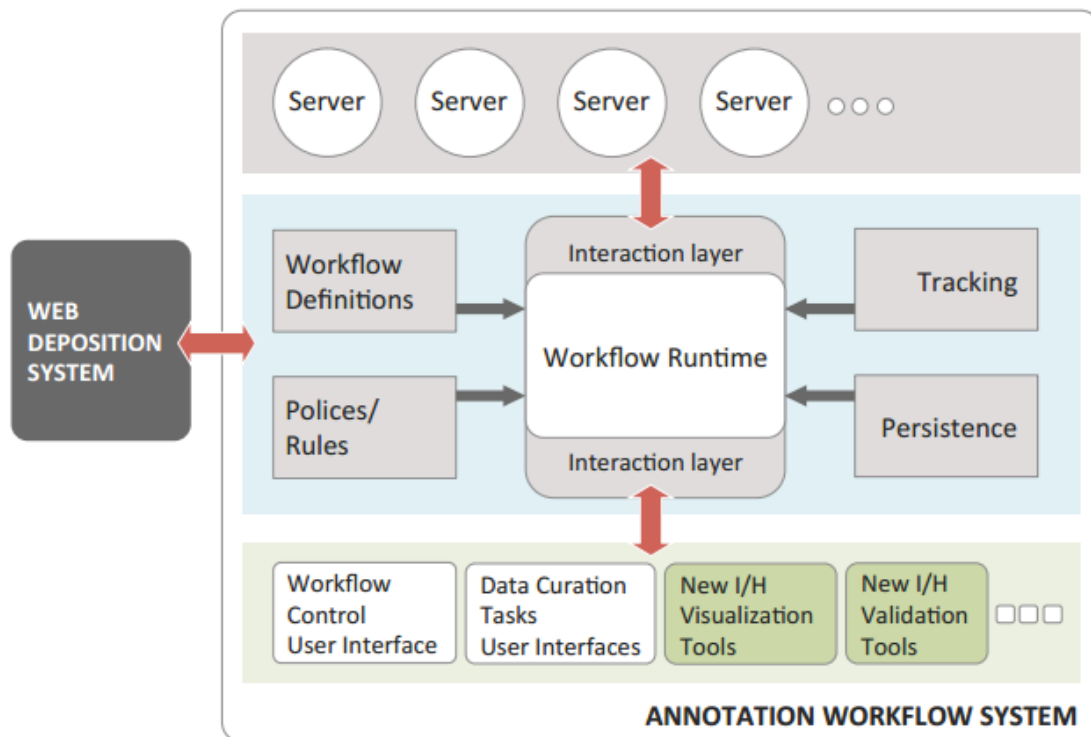


Figure 6. Components of the extensible wwPDB workflow system. It consists of the workflow runtime execution environment, workflow control and curation-task-specific user interfaces, and the supporting compute server infrastructure. The proposed validation and visualization tools for I/H models are highlighted.

Recap

- **Validation is a fundamental part of modeling**
 - Reduce probability of publishing errors
 - Assessment of the quality of the model and data
- **Methods for validating integrative models are under development and not exhaustive**
 - Guide using recent examples
 - Watch for future developments / pipelines in IMP

Future of IMP

- **IMP is under heavy development**
 - 2017 reformulation of the python interface, PMI
 - Check www.integrativemodeling.org
 - **Addition of new experimental methods**
 - Second Harmonic Generation
 - Hydrogen/Deuterium Exchange
 - Fiber Diffraction
 - ???
- **Integration with ChimeraX**
- **Collaboration pushes IMP forward**
 - What interesting problems of yours need solving?